

# High-Q<sup>™</sup>-Spin-Column Fungal Genomic DNA Purification Kit

## Ordering info

TBK0254, 3 reactions (sample)

TBK0255, 50 reactions

# Description

High-Q<sup>™</sup>-Spin-Column Fungal Genomic DNA Purification Kit is an easy silica-membrane-based system for DNA purification from a wide variety of fungal species. An optimized lysis buffer guarantees a good yield while the use of High-Q<sup>™</sup> Spin Columns allow a good quality DNA, suitable for downstream applications.

#### **Features**

- Easy and Fast protocol.
- Typical yields are 2- 50 μg of genomic DNA.
- No organic extraction, DNA is isolated without phenol, chloroform or ethanol precipitation.
- High DNA purity, the isolated DNA is ready to use for sensitive downstream molecular biology applications.
- Isolation of DNA from a wide range of samples.

# **Applications**

- Purification of DNA from fungal samples.
- DNA obtained is suitable for downstream molecular biology applications such as PCR, enzymatic digestion for cloning or Southern, genotyping, etc.

#### **Quality Control**

DNA purified is checked by: integrity (agarose gel electrophoresis), quantity and quality (A260/280= 1.8  $\pm$  0.2).

TBK0256, 200 reactions

TBK0257, 250 reactions

#### **Kit Components**

| Components                                   | TBK0255             | TBK0256            | TBK0257                |
|--|---------------------|--------------------|------------------------|
| High-Q™ Spin Column<br>with Collection Tubes | 50                  | 200                | 250                    |
| BFL1 Buffer                                  | 25 mL               | 85 mL              | 110 mL                 |
| BFL1 Additive                                | 3 mL                | 10 mL              | 12 mL                  |
| BFL2 Buffer                                  | 1.5 mL <sup>a</sup> | 5 mL <sup>b</sup>  | 6 mL <sup>c</sup>      |
| BFL3 Buffer                                  | 7 mL <sup>d</sup>   | 28 mL <sup>e</sup> | 35 mL <sup>f</sup>     |
| BFL4 Buffer                                  | 12 mL <sup>g</sup>  | 45 mL <sup>h</sup> | 2 x 30 mL <sup>i</sup> |
| Elution Buffer                               | 15 mL               | 25 mL              | 25 mL                  |
| Proteinase K                                 | 1.1 mL*             | 3 x 30 mg**        | 4 x 30 mg**            |
| Proteinase K Buffer                          | -                   | 3 x 1.5 mL         | 4 x 1.5 mL             |

Order Info Kit Components: High-Q™ Spin Columns (TBM0010) | BFL1 Buffer (TBB0547) | BFL1 Additive (TBB0548) | BFL2 Buffer (TBB0549) | BFL3 Buffer (TBB0550) | BFL4 Buffer (TBB0551) | Elution Buffer (TBB0510) | Proteinase K 20 mg/ mL (TBZ0308)\* | Proteinase K 30 mg (TBZ0305)\*\* | Proteinase K Buffer (TBB0546).

## ¡Components for samples are ready to use!

## Before its use:

- <sup>a</sup> Add 13.5 mL isopropanol and mix well.
- <sup>b</sup> Add 45 mL isopropanol and mix well.
- <sup>c</sup> Add 54 mL isopropanol and mix well.
- <sup>d</sup> Add 21 mL absolute ethanol and mix well.
- <sup>e</sup> Add 84 mL absolute ethanol and mix well.
- $^{\rm f}$  Add 105 mL absolute ethanol and mix well.
- $^{\rm g}\,\text{Add}$  48 mL absolute ethanol and mix well.
- <sup>h</sup> Add 180 mL absolute ethanol and mix well.
- <sup>i</sup> Add 120 mL absolute ethanol to each bottle. Mix well.

# Storage

Store the kit at 25°C and Proteinase K at -20°C.

#### Material required (not supplied)

- 1.5 mL Tubes.
- Ethanol (CAS 64-17-5).
- Isopropanol (CAS 67-63-0).

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#### **PROTOCOL**

- 1. Grind up to 100 mg of fungal mycelia in liquid nitrogen using a mortar and a pestle. With a freeze spatula, collect the powder into a 1.5mL tube. Commercially available equipment for homogenization can also be used.
- 2. Add 400 μL BFL1 Buffer and 40 μL BFL1 Additive. Vortex briefly.
- 3. Add 20  $\mu$ L Proteinase K (20 mg/ mL). Mix well. Incubate at 65°C, 30 minutes. Mix 2-3 times by inversion during incubation.
- **4.** Centrifuge at 13,000 g for 5 minutes. Transfer the supernatant to a fresh tube.
- 5. [Optional] Add 10 μL RNase-A (20 μg/ μL) and incubate for 5 minutes at room temperature.
- 6. Add 200 μL BFL2 Buffer and mix by inversion. Check isopropanol has been added to BFL2 Buffer (✓).
- 7. Transfer up 700 μL mixture to a High-Q<sup>™</sup> Spin Column placed into a Collection Tube. Incubate 2 minutes at room temperature.
- **8.** Centrifuge at 10,000 g for 1 minute. Remove the flow-through and place back the High-Q<sup>™</sup> Spin Column into a Collection Tube. Repeat steps 7 and 8 with the remaining mixture.
- 9. Add 500 µL BFL3 Buffer.
  Check absolute ethanol has been added to BFL3 Buffer ( ✓).
- **10.** Centrifuge at 10,000 g for 1 minute. Discard the flow-through.
- 11. Place back the High-Q<sup>™</sup> Spin Column into the Collection Tube and add 500 µL BFL4 Buffer. Repeat this step one more time.
  Check absolute ethanol has been added to BFL4 Buffer ( ✓).
- **12.** To dry High-Q<sup>™</sup> Spin Column and eliminate residual ethanol, centrifuge again at 10,000 g for 1 minute.
- **13.** Place the High-Q<sup>™</sup> Spin Column into a clean 1.5 mL Tube.
- 14. Add 50-100  $\mu$ L prewarmed Elution Buffer or Water (Molecular Biology Grade). Prewarm Elution Buffer or Water at 70°C.
- **15.** Incubate at room temperature, 2 minutes.
- **16.** Centrifuge at 10,000 g for 1 minute to elute purified DNA.
- **17.** Check DNA quantity by spectrophotometry and quality on agarose electrophoresis gel.
- **18.** Store DNA at -20°C.

